

PCTWORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

| | | | | | | | | | | | |
|---|--------------------------|--|-------------------------|----|---------|-------------------------|----|---------|--------------------------|----|--|
| (51) International Patent Classification ⁵ : C12N 15/12, A61K 37/02 G01N 33/50, 33/53 | A1 | (11) International Publication Number: WO 94/03600 (43) International Publication Date: 17 February 1994 (17.02.94) | | | | | | | | | |
| (21) International Application Number: PCT/US93/07189 (22) International Filing Date: 29 July 1993 (29.07.93) (30) Priority data: <table border="0"><tr><td>923,780</td><td>31 July 1992 (31.07.92)</td><td>US</td></tr><tr><td>029,335</td><td>4 March 1993 (04.03.93)</td><td>US</td></tr><tr><td>040,510</td><td>31 March 1993 (31.03.93)</td><td>US</td></tr></table> (71) Applicant: CREATIVE BIOMOLECULES, INC. [US/ US]; 45 South Street, Hopkinton, MA 01748 (US). (72) Inventors: JONES, William, K. ; 35 Saint Paul Street, Brookline, MA 02116 (US). TUCKER, Ronald, F. ; 132 Robert Road, Holliston, MA 01746 (US). RUEGER, David, C. ; 19 Downey Street, Hopkinton, MA 01748 (US). OPPERMAN, Hermann ; 25 Summer Hill Road, Medway, MA 02053 (US). OZKAYNAK, Engin ; 44 Purdue Drive, Milford, MA 01757 (US). KUBERA- SAMPATH, Thangavel ; Six Spring Street, Medway, MA 02053 (US). | | 923,780 | 31 July 1992 (31.07.92) | US | 029,335 | 4 March 1993 (04.03.93) | US | 040,510 | 31 March 1993 (31.03.93) | US | (74) Agent: KELLEY, Robin, D. ; Testa, Hurwitz & Thibault, Exchange Place, 53 State Street, Boston, MA 02109 (US). (81) Designated States: AT, AU, BB, BG, BR, CA, CH, CZ, DE, DK, ES, FI, GB, HU, JP, KP, KR, LK, LU, MG, MN, MW, NL, NO, NZ, PL, PT, RO, RU, SD, SE, SK, UA, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the</i> <i>claims and to be republished in the event of the receipt of</i> <i>amendments.</i> |
| 923,780 | 31 July 1992 (31.07.92) | US | | | | | | | | | |
| 029,335 | 4 March 1993 (04.03.93) | US | | | | | | | | | |
| 040,510 | 31 March 1993 (31.03.93) | US | | | | | | | | | |
| (54) Title: MORPHOGENIC PROTEIN SOLUBLE COMPLEX AND COMPOSITION THEREOF (57) Abstract Disclosed are compositions of morphogenic proteins constituting soluble forms of these proteins, antibodies that distinguish between soluble and mature forms, and method for producing these morphogenic proteins and antibodies. | | | | | | | | | | | |

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

| | | | | | |
|----|--------------------------|----|--|----|--------------------------|
| AT | Austria | FR | France | MR | Mauritania |
| AU | Australia | GA | Gabon | MW | Malawi |
| BB | Barbados | GB | United Kingdom | NE | Niger |
| BE | Belgium | CN | Guinea | NL | Netherlands |
| BF | Burkina Faso | GR | Greece | NO | Norway |
| BG | Bulgaria | HU | Hungary | NZ | New Zealand |
| BJ | Benin | IE | Ireland | PL | Poland |
| BR | Brazil | IT | Italy | PT | Portugal |
| BY | Belarus | JP | Japan | RO | Romania |
| CA | Canada | KP | Democratic People's Republic of Korea | RU | Russian Federation |
| CF | Central African Republic | KR | Republic of Korea | SD | Sudan |
| CG | Congo | KZ | Kazakhstan | SE | Sweden |
| CH | Switzerland | LJ | Liechtenstein | SI | Slovenia |
| CI | Côte d'Ivoire | LK | Sri Lanka | SK | Slovak Republic |
| CM | Cameroon | LU | Luxembourg | SN | Senegal |
| CN | China | LV | Latvia | TD | Chad |
| CS | Czechoslovakia | MC | Monaco | TG | Togo |
| CZ | Czech Republic | MG | Madagascar | UA | Ukraine |
| DE | Germany | ML | Mali | US | United States of America |
| DK | Denmark | MN | Mongolia | UZ | Uzbekistan |
| ES | Spain | | | VN | Viet Nam |
| FI | Finland | | | | |

- 1 -

MORPHOGENIC PROTEIN SOLUBLE COMPLEX AND COMPOSITION THEREOF.Field of the Invention

The present invention relates generally to
5 morphogenic proteins and, more particularly, to
compositions having improved solubility in aqueous
solvents.

Background of the Invention

10 Morphogenic proteins ("morphogens") are well known
and described in the art. See, for example, U.S. Pat.
Nos. 4, 968,590; 5,011,691; 5,018,753; PCT US92/01968 and
PCT US92/07432; as well as various articles published in
the scientific literature, including Ozkaynak et al.
15 (1992) J.Biol. Chem. 267:25220-25227 and Ozkaynak et al.
(1991) Biochem. Biophys. Res. Comm. 179:116-123. The
art has described how to isolate morphogenic proteins
from bone, how to identify genes encoding these proteins
and how to express them using recombinant DNA technology.
20 The morphogenic proteins are capable of inducing
endochondral bone formation and other tissue formation in
a mammal when they are properly folded, dimerized and
disulfide bonded to produce a dimeric species having the
appropriate three dimensional conformation. The proteins
25 have utility in therapeutic applications, either by
direct or systemic administration. Where bone induction
is desired, for example, the morphogen typically is
provided to the desired site for bone formation in a
mammal in association with a suitable matrix having the
30 appropriate conformation to allow the infiltration,
proliferation and differentiation of migrating progenitor
cells. The morphogenic protein adsorbed to the surfaces

- 2 -

of a suitable matrix is generally referred to in the art as an osteogenic device. The proteins can be isolated from bone or, preferably, the gene encoding the protein is produced recombinantly in a suitable host cell.

5

The morphogen precursor polypeptide chains share a common structural motif, including a N-terminal signal sequence and pro region, both of which are cleaved to produce a mature sequence, capable of disulfide bonding and comprising an N-terminal extension and a C-terminal domain whose amino acid sequence is highly conserved among members of the family. In their mature dimeric forms, the morphogens typically are fairly insoluble under physiological conditions. Increasing the solubility of these proteins has significant medical utility as it would enhance systemic administration of morphogens as therapeutics. Various carrier proteins, including serum albumin and casein are known to increase the solubility of morphogens (see, for example, PCT US92/07432). PCT US92/05309 (WO 93/00050) discusses the use of various solubilizing agents, including various amino acids and methyl esters thereof, as well as guanidine, sodium chloride and heparin, to increase the solubility of mature dimeric BMP2.

25

Improved methods for the recombinant expression of morphogenic proteins is an ongoing effort in the art. It is an object of this invention to provide an improvement in the methods for producing and purifying morphogenic proteins having high specific activity, and for formulating compositions and osteogenic devices comprising these proteins. Another object is to provide soluble forms of morphogenic proteins consisting essentially of amino acid sequences derived from

30

- 3 -

morphogenic proteins. Another object is to provide formulations which stabilize the soluble complex of morphogenic proteins. Still another object is to provide means for distinguishing between soluble forms of the protein and the mature morphogenic species, to provide means for quantitating the amounts of these proteins in a fluid, including a body fluid, such as serum, cerebro-sprinal fluid or peritoneal fluid, and to provide polyclonal and monoclonal antibodies capable of distinguishing between these various species.

Another object is to provide antibodies and biological diagnostic assays for monitoring the concentration of morphogens and endogenous anti-morphogen antibodies present in a body fluid and to provide kits and assays for detecting fluctuations in the concentrations of these proteins in a body fluid. U.S. Patent No. 4,857,456 and Urist et al. (1984) Proc. Soc. Exp. Biol. Med. 176:472-475 describe a serum assay for detecting a protein purported to be a bone morphogenetic protein. The protein is not a member of the morphogen family of proteins described herein, differing in molecular weight, structural characteristics and solubility from these proteins.

25

Summary of the Invention

It now has been discovered that morphogenic protein secreted into cultured medium from mammalian cells contains as a significant fraction of the secreted protein a soluble form of the protein, and that this soluble form comprises the mature dimeric species, including truncated forms thereof, noncovalently associated with at least one, and preferably two pro domains. It further has been discovered that antibodies

- 4 -

can be used to discriminate between these two forms of the protein. These antibodies may be used as part of a purification scheme to selectively isolate the mature or the soluble form of morphogenic protein, as well as to
5 quantitate the amount of mature and soluble forms produced. These antibodies also may be used as part of diagnostic treatments to monitor the concentration of morphogenic proteins in solution in a body and to detect fluctuations in the concentration of the proteins in
10 their various forms. The antibodies and proteins also may be used in diagnostic assays to detect and monitor concentrations of endogenous anti-morphogen antibodies to the various forms of these proteins in the body.

15 An important embodiment of the invention is a dimeric protein comprising a pair of polypeptide subunits associated to define a dimeric structure having morphogenic activity. As defined herein and in parent, related applications, morphogens generally are capable
20 of all of the following biological functions in a morphogenically permissive environment: stimulating proliferation of progenitor cells; stimulating the differentiation of progenitor cells; stimulating the proliferation of differentiated cells; and supporting the
25 growth and maintenance of differentiated cells.

Each of the subunits of the dimeric morphogenic protein comprises at least the 100 amino acid peptide sequence having the pattern of seven or more cysteine
30 residues characteristic of the morphogen family. Preferably, at least one of the subunits comprises the mature form of a subunit of a member of the morphogen family, or an allelic, species, chimeric or other sequence variant thereof, noncovalently complexed with a

- 5 -

peptide comprising part or all of a pro region of a member of the morphogen family, or an allelic, species, chimeric or other sequence variant thereof. The pair of subunits and one or, preferably, two pro region peptides, together form a complex which is more soluble in aqueous solvents than the uncomplexed pair of subunits.

Preferably, both subunits comprise a mature form of a subunit of a member of the morphogen family or an allelic, species, chimeric or other sequence variant thereof, and both subunits are noncovalently complexed with a peptide comprising a pro region, or a fragment thereof. Most preferably, each subunit is the mature form of human OP-1, or a species, allelic or other sequence variant thereof, and the pro region peptide is the entire or partial sequence of the pro region of human OP-1, or a species, allelic, chimeric or other sequence variant thereof. Currently, preferred pro regions are full length forms of the pro region. Pro region fragments preferably include the first 18 amino acids of the pro sequence. Other useful pro region fragments are truncated sequences of the intact pro region sequence, the truncation occurring at the proteolytic cleavage site Arg-Xaa-Xaa-Arg. As will be appreciated by those having ordinary skill in the art, useful sequences encoding the pro region may be obtained from genetic sequences encoding known morphogens. Alternatively, chimeric pro regions can be constructed from the sequences of one or more known morphogens. Still another option is to create a synthetic sequence variant of one or more known pro region sequences.

- 6 -

As used herein, the mature form of a morphogen protein subunit includes the intact C-terminal domain and intact or truncated forms of the N-terminal extensions. For example, useful mature forms of OP-1 include dimeric species defined by residues 293-431 of Seq ID No. 1, as well as truncated sequences thereof, including sequences defined by residues 300-431, 313-431, 315-431, 316-431 and 318-431. Note that this last sequence retains only about the last 10 residues of the N-terminal extension sequence. Fig. 2 presents the N-terminal extensions for a number of preferred morphogen sequences. Canonical Arg-Xaa-Xaa-Arg cleavage sites where truncation may occur are boxed or underlined in the figure. As will be appreciated by those having ordinary skill in the art, mature dimeric species may include subunit combinations having different N-terminal truncations.

Other soluble forms of morphogens include dimers of the uncleaved pro forms of these proteins (see below), as well as "hemi-dimers" wherein one subunit of the dimer is an uncleaved pro form of the protein, and the other subunit comprises the mature form of the protein, including truncated forms thereof, preferably noncovalently associated with a cleaved pro domain.

The soluble proteins of this invention also are useful in the formation of therapeutic compositions for administration to a mammal, particularly a human, and for the development of biological assays for monitoring the concentration of these proteins and endogenous antibodies to these proteins in cell samples and body fluids, including, but not limited to, serum, cerebrospinal fluid and peritoneal fluid.

- 7 -

The foregoing and other objects, features and advantages of the present invention will be made more apparent from the following detailed description of the invention.

5

Brief Description of the Drawings

Fig. 1 is a schematic representation of a morphogen polypeptide chain as expressed from a nucleic acid
10 encoding the sequence, wherein the cross-hatched region represents the signal sequence; the stippled region represents the pro domain; the hatched region represents the N-terminus ("N-terminal extension") of the mature protein sequence; and the open region represents the
15 C-terminal region of the mature protein sequence defining the conserved seven cysteine domain, the conserved cysteines being indicated by vertical hatched lines;

Fig.2 lists the sequences of the N-terminal
20 extensions of the mature forms of various morphogens; and

Fig. 3 is a gel filtration column elution profile of a soluble morphogen (OP-1) produced and purified from a mammalian cell culture by IMAC, S-Sepharose and S-200HR
25 chromatography in TBS (Tris-buffered saline), wherein V_0 is the void volume, ADH is alcohol dehydrogenase (MW 150 kDa), BSA is bovine serum albumin (MW 67 kDa), CA is carbonic anhydrase (MW 29kDa) and CytC is cytochrome C (MW 12.5 kDa).

30

- 8 -

Detailed Description

A soluble form of morphogenic proteins now has been discovered wherein the proteins consist essentially of the amino acid sequence of the protein. The soluble form is a non-covalently associated complex comprising the pro domain or a fragment thereof, noncovalently associated or complexed with a dimeric protein species having morphogenic activity, each polypeptide of the dimer having less than 200 amino acids and comprising at least the C-terminal six, and preferably seven cysteine skeleton defined by residues 330-431 and 335-431, respectively, of Seq. ID No. 1. Preferably, the polypeptide chains of the dimeric species comprise the mature forms of these sequences, or truncated forms thereof. Preferred truncated forms comprise the intact C-terminal domain and at least 10 amino acids of the N-terminal extension sequence. The soluble forms of these morphogenic proteins may be isolated from cultured cell medium, a mammalian body fluid, or may be formulated in vitro.

In vivo, under physiological conditions, the pro domain may serve to enhance the transportability of the proteins, and/or to protect the proteins from proteases and scavenger molecules, including antibodies. The pro domains also may aid in targeting the proteins to a particular tissue and/or to present the morphogen to a morphogen cell surface receptor by interaction with a co-receptor molecule. The isolated proteins may be used

- 9 -

in therapeutic formulations, particularly for oral or parenteral administration, and in the development of diagnostic and other tissue evaluating kits and assays to monitor the level of endogenous morphogens and endogenous anti-morphogen antibodies.

Detailed descriptions of the utility of these morphogens in therapies to regenerate lost or damaged tissues and/or to inhibit the tissue destructive effects of tissue disorders or diseases, are provided in international applications US92/01968 (WO92/15323); US92/07358 (WO93/04692) and US92/07432 (WO93/05751) the disclosures of which are incorporated herein by reference. Morphogens, including the soluble morphogen complexes of this invention, are envisioned to have particular utility as part of therapies for regenerating lost or damaged bone, dentin, periodontal, liver, cardiac, lung and nerve tissue, as well as for protecting these tissues from the tissue destructive effects associated with an immunological response. The proteins also are anticipated to provide a tissue protective effect in the treatment of metabolic bone disorders, such as osteoporosis, osteomalacia and osteosarcoma; in the treatment of liver disorders, including cirrhosis, hepatitis, alcohol liver disease and hepatic encephalopathy; and in the treatment or prevention of ischemia reperfusion-associated tissue damage, particularly to nerve or cardiac tissue.

30

- 10 -

Presented below are detailed descriptions of useful soluble morphogen complexes of this invention, as well as how to make and use them.

5 I. Useful Soluble Morphogen Complexes -
Protein Considerations

Among the morphogens useful in this invention are proteins originally identified as osteogenic proteins,
10 such as the OP-1, OP-2 and CBMP2 proteins, as well as amino acid sequence-related proteins such as DPP (from *Drosophila*), Vgl (from *Xenopus*), Vgr-1 (from mouse, see U.S. 5,011,691 to Oppermann et al.), GDF-1 (from mouse, see Lee (1991) PNAS 88:4250-4254), 60A protein (from
15 *Drosophila*, Seq. ID No. 24, see Wharton et al. (1991) PNAS 88:9214-9218), and the recently identified OP-3.

The members of this family, which are a subclass of the TGF- β super-family of proteins, share characteristic
20 structural features, represented schematically in Fig. 1, as well as substantial amino acid sequence homology in their C-terminal domains, including a conserved seven cysteine structure. As illustrated in the figure, the proteins are translated as a precursor polypeptide
25 sequence 10, having an N-terminal signal peptide sequence 12, (the "pre pro" region, indicated in the figure by cross-hatching), typically less than about 30 residues, followed by a "pro" region 14, indicated in the figure by stippling, and which is cleaved to yield the mature
30 sequence 16. The mature sequence comprises both the conserved C-terminal seven cysteine domain 20, and an N-terminal sequence 18, referred to herein as an N-terminal xtension, and which varies significantly in sequence between the various morphogens. Cysteines are

- 11 -

represented in the figure by vertical hatched lines 22.
The polypeptide chains dimerize and these dimers
typically are stabilized by at least one interchain
disulfide bond linking the two polypeptide chain
5 subunits.

The signal peptide is cleaved rapidly upon
translation, at a cleavage site that can be predicted in
a given sequence using the method of Von Heijne ((1986)
10 Nucleic Acids Research 14:4683-4691.) The "pro" form of
the protein subunit, 24, in Fig. 1, includes both the pro
domain and the mature domain, peptide bonded together.
Typically, this pro form is cleaved while the protein is
still within the cell, and the pro domain remains
15 noncovalently associated with the mature form of the
subunit to form a soluble species that appears to be the
primary form secreted from cultured mammalian cells.
Typically, previous purification techniques utilized
denaturing conditions that disassociated the complex.

20 Other soluble forms of morphogens secreted from
mammalian cells include dimers of the pro forms of these
proteins, wherein the pro region is not cleaved from the
mature domain, and "hemi-dimers", wherein one subunit
25 comprises a pro form of the polypeptide chain subunit and
the other subunit comprises the cleaved mature form of
the polypeptide chain subunit (including truncated forms
thereof), preferably noncovalently associated with a
cleaved pro domain.

30 The isolated pro domain typically has a substantial
hydrophobic character, as determined both by analysis of
the sequence and by characterization of its properties in
solution. The isolated pro regions alone typically are

- 12 -

not significantly soluble in aqueous solutions, and require the presence of denaturants, e.g., detergents, urea, guanidine HCl, and the like, and/or one or more carrier proteins. Accordingly, without being limited to
5 any given theory, the non-covalent association of the cleaved pro region with the mature morphogen dimeric species likely involves interaction of a hydrophobic portion of the pro region with a corresponding hydrophobic region on the dimeric species, the
10 interaction of which effectively protects or "hides" an otherwise exposed hydrophobic region of the mature dimer from exposure to aqueous environments, enhancing the affinity of the mature dimer species for aqueous solutions.

15

Morphogens comprise a subfamily of proteins within the TGF- β superfamily of structurally related proteins. Like the morphogens described herein, TGF- β also has a pro region which associates non-covalently with the
20 mature TGF- β protein form. However, unlike the morphogens, the TGF- β pro region contains numerous cysteines and forms disulfide bonds with a specific binding protein. The TGF- β 1 pro domain also is phosphorylated at one or more mannose residues, while the
25 morphogen pro regions typically are not.

Useful pro domains include the full length pro regions described below, as well as various truncated forms hereof, particularly truncated forms cleaved at
30 proteolytic Arg-Xaa-Xaa-Arg cleavage sites. For example, in OP-1, possible pro sequences include sequences defined by residues 30-292 (full length form); 48-292; and 158-292. Soluble OP-1 complex stability is enhanced when the pro region comprises the full length form rather than

- 13 -

a truncated form, such as the 48-292 truncated form, in that residues 30-47 show sequence homology to the N-terminal portions of other morphogens, and are believed to have particular utility in enhancing complex stability for all morphogens. Accordingly, currently preferred pro sequences are those encoding the full length form of the pro region for a given morphogen (see below). Other pro sequences contemplated to have utility include biosynthetic pro sequences, particularly those that incorporate a sequence derived from the N-terminal portion of one or more morphogen pro sequences.

Table I, below, describes the various preferred morphogens identified to date, including their nomenclature as used herein, the sequences defining the various regions of the subunit sequences, their Seq. ID references, and publication sources for their nucleic acid and amino acid sequences. The disclosure of these publications is incorporated herein by reference. The mature protein sequences defined are the longest anticipated forms of these sequences. As described above, shorter, truncated forms of these sequences also are contemplated. Preferably, truncated mature sequences include at least 10 amino acids of the N-terminal extension. Fig. 2 lists the N-terminal extensions for a number of the preferred morphogen sequences described below. Arg-Xaa-Xaa-Arg cleavage sites that may yield truncated sequences of the mature subunit form are boxed or underlined in the figure.

30

- 14 -

TABLE I

| | | |
|----|--------|---|
| 5 | "OP-1" | Refers generically to the group of morphogenically active proteins expressed from part or all of a DNA sequence encoding OP-1 protein, including allelic and species variants thereof, e.g., human OP-1 ("hOP-1"), or mouse OP-1 ("mOP-1"). |
| 10 | | The cDNA sequences and the amino acids encoding the full length proteins are provided in Seq. Id Nos. 1 and 2 (hOP1) and Seq. ID Nos. 3 and 4 (mOP1.) |
| 15 | | The mature proteins are defined by residues 293-431 (hOP1) and 292-430 (mOP1), wherein the conserved seven cysteine skeleton is defined by residues 330-431 and 329-430, respectively, and the N-terminal extensions are defined by residues 293-329 and 292-329, respectively. |
| 20 | | The "pro" regions of the proteins, cleaved to yield the mature, morphogenically active proteins, are defined essentially by residues 30-292 (hOP1) and residues 30-291 (mOP1). |
| 25 | | |
| 30 | "OP-2" | refers generically to the group of active proteins expressed from part or all of a DNA sequence encoding OP-2 protein, including allelic and species variants thereof, e.g., human OP-2 ("hOP-2") or mouse OP-2 ("mOP-2"). |
| | | The full length proteins are provided in Seq. ID Nos. 5 and 6 (hOP2) and Seq. ID Nos. 7 and 8 (mOP2.) |
| | | The mature proteins are defined |

- 15 -

essentially by residues 264-402 (hOP2) and
261-399 (mOP2), wherein the conserved
seven cysteine skeleton is defined by
residues 301-402 and 298-399,
5 respectively, and the N-terminal
extensions are defined by residues 264-300
and 261-297, respectively. The "pro"
regions of the proteins, cleaved to yield
the mature, morphogenically active
10 proteins likely are defined essentially by
residues 18-263 (hOP2) and residues 18-260
(mOP2). (Another cleavage site also
occurs 21 residues upstream for both OP-2
proteins.)

15 "OP-3" refers generically to the group of active
proteins expressed from part or all of a
DNA sequence encoding OP-3 protein,
including allelic and species variants
20 thereof, e.g., mouse OP-3 ("mOP-3".) The
full length protein is provided in Seq. ID
No. 9. The mature protein is defined
essentially by residues 261-399 or
264-399, wherein the conserved seven
25 cysteine skeleton is defined by residues
298-399 and the N-terminal extension is
defined by residues 264-297 or 261-297.
The "pro" region of the protein, cleaved
to yield the mature, morphogenically
30 active proteins likely is defined
essentially by residues 20-262.

- 16 -

"BMP2/BMP4" refers to protein sequences encoded by the human BMP2 and BMP4 genes. The amino acid sequence for the full length proteins, referred to in the literature as BMP2A and BMP2B, or BMP2 and BMP4, appear in Seq. ID Nos. 10 and 11, respectively, and in Wozney, et al. (1988) Science 242:1528-1534. The pro domain for BMP2 (BMP2A) likely includes residues 25-248 or 25-282; the mature protein, residues 249-396 or 283-396, of which residues 249-296/283-296 define the N-terminal extension and 295-396 define the C-terminal domain. The pro domain for BMP4 (BMP2B) likely includes residues 25-256 or 25-292; the mature protein, residues 257-408 or 293-408, of which 257-307/293-307 define the N-terminal extension, and 308-408 define the C-terminal domain.

"DPP" refers to protein sequences encoded by the Drosophila DPP gene. The amino acid sequence for the full length protein, including the mature form and the pro region, appears in Seq.ID No. 12 and in Padgett, et al (1987) Nature 325: 81-84. The pro domain likely extends from the signal peptide cleavage site to residue 456; the mature protein likely is defined by residues 457-588, where residues 457-586 define the N-terminal extension and 487-588 define the C-terminal domain.

- 17 -

- "Vgl" refers to protein sequences encoded by the Xenopus Vgl gene. The amino acid sequence for the full length protein, including the mature form and the pro region, appears in Seq.ID No. 13 and in Weeks (1987) Cell 51: 861-867. The pro domain likely extends from the signal peptide cleavage site to residue 246; the mature protein likely is defined by residues 247-360, where residues 247-258 define the N-terminal extension, and residues 259-360 define the C-terminal domain.
- "Vgr-1" refers to protein sequences encoded by the murine Vgr-1 gene. The amino acid sequence for the full length protein, including the mature form and the pro region, appears in Seq. ID No. 14 and in Lyons, et al, (1989) PNAS 86: 4554-4558. The pro domain likely extends from the signal peptide cleavage site to residue 299; the mature protein likely is defined by residues 300-438, where residues 300-336 define the N-terminal extension and residues 337-438 define the C-terminus.
- "GDF-1" refers to protein sequences encoded by the human GDF-1 gene. The cDNA and encoded amino sequence for the full length protein is provided in Seq. ID. No. 15 and Lee (1991) PNAS 88:4250-4254. The pro domain

- 18 -

- 5 likely extends from the signal peptide cleavage site to residue 214; the mature protein likely is defined by residues 215-372, where residues 215-256 define the N-terminal extension and residues 257-372 define the C-terminus.
- 10 "60A" refers to protein sequences encoded by the *Drosophila* 60A gene. The amino acid sequence for the full length protein appears in Seq. ID No. 16 and in Wharton et al. (1991) PNAS 88:9214-9218) The pro domain likely extends from the signal peptide cleavage site to residue 324; the
- 15 mature protein likely is defined by residues 325-455, wherein residues 325-353 define the N-terminal extension and residues 354-455 define the C-terminus.
- 20 "BMP3" refers to protein sequences encoded by the human BMP3 gene. The amino acid sequence for the full length protein, including the mature form and the pro region, appears in Seq.ID No. 17 and in Wozney et al. (1988)
- 25 Science 242: 1528-1534. The pro domain likely extends from the signal peptide cleavage site to residue 290; the mature protein likely is defined by residues 291-472, wherein residues 291-370 define the
- 30 N-terminal extension and residues 371-472 define the C-terminus.

- 19 -

- "BMP5" refers to protein sequences encoded by the human BMP5 gene. The amino acid sequence for the full length protein, including the mature form and the pro region, appears in Seq.ID No. 18 and in Celeste, et al. (1990) PNAS 87: 9843-9847. The pro domain likely extends from the signal peptide cleavage site to residue 316; the mature protein likely is defined by residues 317-454, where residues 317-352 define the N-terminus and residues 352-454 define the C-terminus.
- "BMP6" refers to protein sequences encoded by the human BMP6 gene. The amino acid sequence for the full length protein, including the mature form and the pro region, appears in Seq. ID No. 16 and in Celeste, et al. (1990) PNAS 87: 9843-5847. The pro domain likely includes extends from the signal peptide cleavage site to residue 374; the mature sequence likely includes residues 375-513, where residues 375-411 define the N-terminus and residues 412-513 define the C-terminus.

Note that the OP-2 and OP-3 proteins have an additional cysteine residue in the C-terminal region (e.g., see residue 338 in these sequences), in addition to the conserved cysteine skeleton in common with the other proteins in this family. The GDF-1 protein has a four amino acid insert within the conserved skeleton

- 20 -

("Gly-Gly-Pro-Pro") but this insert likely does not interfere with the relationship of the cysteines in the folded structure. In addition, the CBMP2 proteins are missing one amino acid residue within the cysteine
5 skeleton.

The dimeric morphogen species are inactive when reduced, but are active as oxidized homodimers and when oxidized in combination with other morphogens of this
10 invention. Thus, as defined herein, a morphogen useful in a soluble morphogen complex is a dimeric protein comprising a pair of polypeptide chains, wherein each polypeptide chain has less than 200 amino acids and comprises at least the C-terminal six, preferably seven
15 cysteine skeleton defined by residues 335-431 of Seq. ID No. 1, including functionally equivalent arrangements of these cysteines (e.g., amino acid insertions or deletions which alter the linear arrangement of the cysteines in the sequence but not
20 their relationship in the folded structure), such that, when the polypeptide chains are folded, the dimeric protein species comprising the pair of polypeptide chains has the appropriate three-dimensional structure, including the appropriate intra- or inter-chain
25 disulfide bonds such that the protein is capable of acting as a morphogen as defined herein. The solubility of these structures is improved when the mature dimeric form of a morphogen, in accordance with the invention, is complexed with at least one, and
30 preferably two, pro domains.

- 21 -

Various generic sequences (Generic Sequence 1-6) defining preferred C-terminal sequences useful in the soluble morphogens of this invention are described in USSN 07/923,780, incorporated herein above by

5 reference. Two currently preferred generic sequences are described below.

Generic Sequence 7 (Seq. ID No. 20) and Generic Sequence 8 (Seq. ID No. 21) disclosed below,
10 accommodate the homologies shared among preferred morphogen protein family members identified to date, including OP-1, OP-2, OP-3, CBMP2A, CBMP2B, BMP3, 60A, DPP, Vgl, BMP5, BMP6, Vrg-1, and GDF-1. The amino acid sequences for these proteins are described herein (see
15 Sequence Listing) and/or in the art, as well as in PCT publication US 92/07358, (WO93/04692), for example. The generic sequences include both the amino acid identity shared by these sequences in the C-terminal domain, defined by the six and seven cysteine skeletons
20 (Generic Sequences 7 and 8, respectively), as well as alternative residues for the variable positions within the sequence. The generic sequences allow for an additional cysteine at position 41 (Generic Sequence 7) or position 46 (Generic Sequence 8), providing an
25 appropriate cysteine skeleton where inter- or intramolecular disulfide bonds can form, and containing certain critical amino acids which influence the tertiary structure of the proteins.

- 22 -

Generic Sequence 7

```

          Leu Xaa Xaa Xaa Phe
            1               5
5      Xaa Xaa Xaa Gly Trp Xaa Xaa Xaa Xaa
              10
      Xaa Xaa Pro Xaa Xaa Xaa Xaa Ala
        15               20
      Xaa Tyr Cys Xaa Gly Xaa Cys Xaa
10              25               30
      Xaa Pro Xaa Xaa Xaa Xaa Xaa
              35
      Xaa Xaa Xaa Asn His Ala Xaa Xaa
        40               45
15     Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
              50
      Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys
        55               60
      Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa
20              65
      Xaa Xaa Xaa Leu Xaa Xaa Xaa
        70               75
      Xaa Xaa Xaa Xaa Val Xaa Leu Xaa
              80
25     Xaa Xaa Xaa Xaa Met Xaa Val Xaa
        85               90
      Xaa Cys Xaa Cys Xaa
              95

```

wherein each Xaa is independently selected from a group
 30 of one or more specified amino acids defined as
 follows: "Res." means "residue" and Xaa at res.2 =
 (Tyr or Lys); Xaa at res.3 = Val or Ile); Xaa at res.4
 = (Ser, Asp or Glu); Xaa at res.6 = (Arg, Gln, Ser, Lys
 or Ala); Xaa at res.7 = (Asp or Glu); Xaa at res.8 =

- 23 -

(Leu, Val or Ile); Xaa at res.11 = (Gln, Leu, Asp, His, Asn or Ser); Xaa at res.12 = (Asp, Arg, Asn or Glu); Xaa at res. 13 = (Trp or Ser); Xaa at res.14 = (Ile or Val); Xaa at res.15 = (Ile or Val); Xaa at res.16 (Ala or Ser); Xaa at res.18 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.19 = (Gly or Ser); Xaa at res.20 = (Tyr or Phe); Xaa at res.21 = (Ala, Ser, Asp, Met, His, Gln, Leu or Gly); Xaa at res.23 = (Tyr, Asn or Phe); Xaa at res.26 = (Glu, His, Tyr, Asp, Gln, Ala or Ser); Xaa at res.28 = (Glu, Lys, Asp, Gln or Ala); Xaa at res.30 = (Ala, Ser, Pro, Gln, Ile or Asn); Xaa at res.31 = (Phe, Leu or Tyr); Xaa at res.33 = (Leu, Val or Met); Xaa at res.34 = (Asn, Asp, Ala, Thr or Pro); Xaa at res.35 = (Ser, Asp, Glu, Leu, Ala or Lys); Xaa at res.36 = (Tyr, Cys, His, Ser or Ile); Xaa at res.37 = (Met, Phe, Gly or Leu); Xaa at res.38 = (Asn, Ser or Lys); Xaa at res.39 = (Ala, Ser, Gly or Pro); Xaa at res.40 = (Thr, Leu or Ser); Xaa at res.44 = (Ile, Val or Thr); Xaa at res.45 = (Val, Leu, Met or Ile); Xaa at res.46 = (Gln or Arg); Xaa at res.47 = (Thr, Ala or Ser); Xaa at res.48 = (Leu or Ile); Xaa at res.49 = (Val or Met); Xaa at res.50 = (His, Asn or Arg); Xaa at res.51 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.52 = (Ile, Met, Asn, Ala, Val, Gly or Leu); Xaa at res.53 = (Asn, Lys, Ala, Glu, Gly or Phe); Xaa at res.54 = (Pro, Ser or Val); Xaa at res.55 = (Glu, Asp, Asn, Gly, Val, Pro or Lys); Xaa at res.56 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser, Gly, Ile or His); Xaa at res.57 = (Val, Ala or Ile); Xaa at res.58 = (Pro or Asp); Xaa at res.59 = (Lys, Leu or Glu); Xaa at res.60 = (Pro, Val or Ala); Xaa at res.63 = (Ala or Val); Xaa at res.65 = (Thr, Ala or Glu); Xaa at res.66 = (Gln, Lys, Arg or Glu); Xaa at res.67 = (Leu, Met or Val); Xaa at res.68 = (Asn, Ser, Asp or Gly); Xaa at

- 24 -

res.69 = (Ala, Pro or Ser); Xaa at res.70 = (Ile, Thr, Val or Leu); Xaa at res.71 = (Ser, Ala or Pro); Xaa at res.72 = (Val, Leu, Met or Ile); Xaa at res.74 = (Tyr or Phe); Xaa at res.75 = (Phe, Tyr, Leu or His); Xaa at res.76 = (Asp, Asn or Leu); Xaa at res.77 = (Asp, Glu, Asn, Arg or Ser); Xaa at res.78 = (Ser, Gln, Asn, Tyr or Asp); Xaa at res.79 = (Ser, Asn, Asp, Glu or Lys); Xaa at res.80 = (Asn, Thr or Lys); Xaa at res.82 = (Ile, Val or Asn); Xaa at res.84 = (Lys or Arg); Xaa at res.85 = (Lys, Asn, Gln, His, Arg or Val); Xaa at res.86 = (Tyr, Glu or His); Xaa at res.87 = (Arg, Gln, Glu or Pro); Xaa at res.88 = (Asn, Glu, Trp or Asp); Xaa at res.90 = (Val, Thr, Ala or Ile); Xaa at res.92 = (Arg, Lys, Val, Asp, Gln or Glu); Xaa at res.93 = (Ala, Gly, Glu or Ser); Xaa at res.95 = (Gly or Ala) and Xaa at res.97 = (His or Arg).

As described above, Generic Sequence 8 (Seq. ID No. 21) includes all of Generic Sequence 7 and in addition includes the following sequence at its N-terminus:

Cys Xaa Xaa Xaa Xaa
1 5

25 Accordingly, beginning with residue 7, each "Xaa"
in Generic Seq. 8 is a specified amino acid defined as
for Generic Seq. 7, with the distinction that each
residue number described for Generic Sequence 7 is
shifted by five in Generic Seq. 8. Thus, "Xaa at res.2
30 =(Tyr or Lys)" in Gen. Seq. 7 refers to Xaa at res. 7

- 25 -

in Generic Seq. 8. In Generic Seq. 8, Xaa at res.2 = (Lys, Arg, Ala or Gln); Xaa at res.3 = (Lys, Arg or Met); Xaa at res.4 = (His, Arg or Gln); and Xaa at res.5 = (Glu, Ser, His, Gly, Arg, Pro, Thr, or Tyr).

5

Accordingly, other useful sequences defining preferred C-terminal sequences are those sharing at least 70% amino acid sequence homology or "similarity", and preferably 80% homology or similarity with any of the sequences incorporated into Generic Seq. 7 and 8 above. These are anticipated to include allelic, species, chimeric and other sequence variants, (e.g., including "muteins" or "mutant proteins"), whether naturally-occurring or biosynthetically produced, as well as novel members of this morphogenic family of proteins. As used herein, "amino acid sequence homology" is understood to mean amino acid sequence similarity, and homologous sequences share identical or similar amino acids, where similar amino acids are conserved amino acids as defined by Dayoff et al., Atlas of Protein Sequence and Structure; vol.5, Suppl.3, pp.345-362 (M.O. Dayoff, ed., Nat'l BioMed. Research Fdn., Washington D.C. 1978.) Thus, a candidate sequence sharing 70% amino acid homology with a reference sequence requires that, following alignment of the candidate sequence with the reference sequence, 70% of the amino acids in the candidate sequence are identical to the corresponding amino acid in the reference sequence, or constitute a conserved amino acid change thereto. "Amino acid sequence identity" is understood to require identical amino acids between two

10

15

20

25

30

- 26 -

aligned sequences. Thus, a candidate sequence sharing 60% amino acid identity with a reference sequence requires that, following alignment of the candidate sequence with the reference sequence, 60% of the amino acids in the candidate sequence are identical to the corresponding amino acid in the reference sequence.

As used herein, all homologies and identities calculated use OP-1 as the reference sequence. Also as used herein, sequences are aligned for homology and identity calculations using the method of Needleman et al. (1970) J.Mol. Biol. 48:443-453 and identities calculated by the Align program (DNASTar, Inc.) In all cases, internal gaps and amino acid insertions in the candidate sequence as aligned are ignored when making the homology/identity calculation.

Also as used herein, "sequence variant" is understood to mean an amino acid sequence variant form of the morphogen protein, wherein the amino acid change or changes in the sequence do not alter significantly the morphogenic activity (e.g., tissue regeneration activity) of the protein, and the variant molecule performs substantially the same function in substantially the same way as the naturally-occurring form of the molecule. Sequence variants may include single or multiple amino acid changes, and are intended to include chimeric sequences as described below. The variants may be naturally-occurring or may be biosynthetically induced by using standard recombinant DNA techniques or chemical protein synthesis methodologies.

- 27 -

The currently most preferred protein sequences useful in soluble morphogen complexes in this invention include those having greater than 60% identity, preferably greater than 65% identity, with the amino acid sequence defining the conserved six cysteine skeleton of hOP1 (e.g., residues 335-431 of Seq. ID No. 5). These most preferred sequences include both allelic and species variants of the OP-1 and OP-2 proteins, including the *Drosophila* 60A protein.

10 Accordingly, in another preferred aspect of the invention, useful morphogens include active proteins comprising species of polypeptide chains having the generic amino acid sequence herein referred to as "OPX", which accommodates the homologies between the

15 various identified species of OP1 and OP2 (Seq. ID No. 22).

In still another preferred aspect of the invention, useful morphogens include active proteins comprising

20 amino acid sequences encoded by nucleic acids that hybridize to DNA or RNA sequences encoding the conserved C-terminal cysteine domain of OP1 or OP2, e.g., defined by nucleotides 1036-1341 and nucleotides 1390-1695 of Seq. ID Nos. 1 and 5, respectively, under

25 stringent hybridization conditions. As used herein, stringent hybridization conditions are defined as hybridization in 40% formamide, 5 X SSPE, 5 X Denhardt's Solution, and 0.1% SDS at 37°C overnight, and washing in 0.1 X SSPE, 0.1% SDS at 50°C.

30 Similarly, in another preferred aspect of the invention, useful pro region peptides include polypeptide chains comprising amino acid sequences encoded by nucleic acids that hybridize to DNA or RNA sequences encoding at least the N-terminal 18 amino

- 28 -

acids of the pro region sequences for any of the sequences listed in Seq. ID Nos. 1-19, under stringent hybridization conditions. Most preferably, the peptides are encoded by nucleic acids that hybridize to
5 the DNA or RNA sequences encoding at least the N-terminal 18 amino acids of the pro region sequences for OP1 or OP2, e.g., nucleotides 136-192 and nucleotides 152-211 of Seq. ID Nos. 1 and 5, respectively.

10

Useful N-terminal extension sequences are listed in Fig. 2 for use with the C-terminal domains described above. Also as described above, the full length N-terminal extensions, or truncated forms thereof, may be
15 used in preferred dimeric species. The mature dimeric species may be produced from intact DNAs, or truncated forms thereof. It also is envisioned as an embodiment of the invention that chimeric morphogen sequences can be used. Thus, DNAs encoding chimeric morphogens may
20 be constructed using part or all of the N-terminal extension from one morphogen and a C-terminal domain derived from one or more other morphogens. These chimeric proteins may be synthesized using standard recombinant DNA methodology and/or automated chemical
25 nucleic acid synthesis methodology well described in the art. Other chimeric morphogens include soluble morphogen complexes where the pro domain is encoded from a DNA sequence corresponding to one or more morphogen pro sequences, and part or all of the mature
30 domain is encoded by DNA derived from one or more

- 29 -

other, different morphogens. These soluble chimerics may be produced from a single synthetic DNA as described below, or, alternatively, may be formulated in vitro from isolated components also as described
5 herein below.

Finally, the morphogen pro domains and/or mature form N-terminal extensions themselves may be useful as tissue targeting sequences. As described above, the
10 morphogen family members share significant sequence homology in their C-terminal active domains. By contrast, the sequences diverge significantly in the sequences which define the pro domain and the N-terminal 39 amino acids of the mature protein.
15 Accordingly, the pro domain and/or N-terminal extension sequence may be morphogen-specific. Accordingly, part or all of these morphogen-specific sequences may serve as tissue targeting sequences for the morphogens described herein. For example, the N-terminal
20 extension and/or pro domains may interact specifically with one or more molecules at the target tissue to direct the morphogen associated with the pro domain to that tissue. Thus, for example, the morphogen-specific sequences of OP-1, BMP2 or BMP4, all of which proteins
25 are found naturally associated with bone tissue (see, for example, US Pat. No. 5,011,691) may be particularly useful sequences when the morphogen complex is to be targeted to bone. Similarly, BMP6 (or Vgr-1) specific sequences may be used when targeting to lung tissue is
30 desired. Alternatively, the morphogen-specific sequences of GDF-1 may be used to target soluble

- 30 -

morphogen complexes to nerve tissue, particularly brain tissue, where GDF-1 appears to be primarily expressed (see, for example, Lee, PNAS, 88:4250-4254 (1991), incorporated herein by reference).

5

II. Recombinant Production of Soluble Morphogen Complexes

Soluble morphogen complexes can be produced from
10 eukaryotic host cells, preferably mammalian cells,
using standard recombinant expression techniques. An
exemplary protocol currently preferred, is provided
below, using a particular vector construct and chinese
hamster ovary (CHO) cell line. Those skilled in the
15 art will appreciate that other expression systems are
contemplated to be useful, including other vectors and
other cell systems, and the invention is not intended
to be limited to soluble morphogenic protein complexes
produced only by the method detailed hereinbelow.
20 Similar results to those described herein have been
observed using recombinant expression systems developed
for COS and BSC cells.

Morphogen DNA encoding the precursor sequence is
25 subcloned into an insertion site of a suitable,
commercially available pUC-type vector (e.g., pUC-19,
ATCC #37254, Rockville, MD), along with a suitable
promoter/enhancer sequences and 3' termination
sequences. Useful DNA sequences include the published
30 sequences encoding these proteins, and/or synthetic
constructs. Currently preferred promoter/enhancer
sequences are the CMV promoter (human cytomegalovirus
major intermediate - early promoter) and the mouse

- 31 -

mammary tumor virus promoter (mMTV) boosted by the rous sarcoma virus LTR enhancer sequence (e.g., from Clontech, Inc., Palo Alto). Expression also may be further enhanced using transactivating enhancer
5 sequences. The plasmid also contains DHFR as an amplifiable marker, under SV40 early promoter control (ATCC #37148). Transfection, cell culturing, gene amplification and protein expression conditions are standard conditions, well known in the art, such as are
10 described, for example in Ausubel et al., ed., Current Protocols in Molecular Biology, John Wiley & Sons, NY (1989). Briefly, transfected cells are cultured in medium containing 0.1-0.5% dialyzed fetal calf serum (FCS) and stably transfected high expression cell lines
15 are obtained by subcloning and evaluated by standard Western or Northern blot. Southern blots also are used to assess the state of integrated sequences and the extent of their copy number amplification.

20 A currently preferred expression vector contains the DHFR gene, under SV40 early promoter control, as both a selection marker and as an inducible gene amplifier. The DNA sequence for DHFR is well characterized in the art, and is available
25 commercially. For example, a suitable vector may be generated from pMAM-neo (Clontech, Inc., Palo Alto, CA) by replacing the neo gene (BamHI digest) with an SphI-BamHI, or a PvuII-BamHI fragment from pSV5-DHFR (ATCC #37148), which contains the DHFR gene under SV40 early
30 promoter control. A BamHI site can be engineered at the SphI or PvuII site using standard techniques (e.g., by linker insertion or site-directed mutagenesis) to allow insertion of the fragment into the vector backbone. The morphogen DNA can be inserted into the

- 32 -

polylinker site downstream of the MMTV-LTR sequence (mouse mammary tumor virus LTR). The CMV promoter sequence then may be inserted into the expression vector (e.g., from pCDM8, Invitrogen, Inc.) The SV40
5 early promoter, which drives DHFR expression, preferably is modified in these vectors to reduce the level of DHFR mRNA produced.

The currently preferred mammalian cell line is a
10 CHO Chinese hamster ovary, cell line, and the preferred procedure for establishing a stable morphogen production cell line with high expression levels comprises transfecting a stable CHO cell line, preferably CHO-DXB11, with the expression vector
15 described above, isolating clones with high morphogen expression levels, and subjecting these clones to cycles of subcloning using a limited dilution method described below to obtain a population of high expression clones. Subcloning preferably is performed
20 in the absence of MTX to identify stable high expression clones which do not require addition of MTX to the growth media for morphogen production.

In the subcloning protocol cells are seeded on ten
25 100mm petri dishes at a cell density of either 50 or 100 cells per plate, with or preferably without MTX in the culture media. After 14 days of growth, clones are isolated using cloning cylinders and standard procedures, and cultured in 24-well plates. Clones
30 then are screened for morphogen expression by Western

- 33 -

immunoblots using standard procedures, and morphogen expression levels compared to parental lines. Cell line stability of high expression subclones then is determined by monitoring morphogen expression levels over multiple cell passages (e.g., four or five passages).

III. Isolation of Soluble morphogen complex from conditioned media or body fluid

10

Morphogens are expressed from mammalian cells as soluble complexes. Typically, however the complex is disassociated during purification, generally by exposure to denaturants often added to the purification solutions, such as detergents, alcohols, organic solvents, chaotropic agents and compounds added to reduce the pH of the solution. Provided below is a currently preferred protocol for purifying the soluble proteins from conditioned media (or, optionally, a body fluid such as serum, cerebro-spinal or peritoneal fluid), under non-denaturing conditions. The method is rapid, reproducible and yields isolated soluble morphogen complexes in substantially pure form.

25 Soluble morphogen complexes can be isolated from conditioned media using a simple, three step chromatographic protocol performed in the absence of denaturants. The protocol involves running the media (or body fluid) over an affinity column, followed by ion exchange and gel filtration chromatographies. The affinity column described below is a Zn-IMAC column. The present protocol has general applicability to the purification of a variety of morphogens, all of which are anticipated to be isolatable using only minor

30

- 34 -

modifications of the protocol described below. An alternative protocol also envisioned to have utility an immunoaffinity column, created using standard procedures and, for example, using antibody specific
5 for a given morphogen pro domain (complexed, for example, to a protein A-conjugated Sepharose column.) Protocols for developing immunoaffinity columns are well described in the art, (see, for example, Guide to Protein Purification, M. Deutscher, ed., Academic
10 Press, San Diego, 1990, particularly sections VII and XI.)

In this experiment OP-1 was expressed in CHO cells as described above. The CHO cell conditioned media
15 containing 0.5% FBS was initially purified using Immobilized Metal-Ion Affinity Chromatography (IMAC). The soluble OP-1 complex from conditioned media binds very selectively to the Zn-IMAC resin and a high concentration of imidazole (50 mM imidazole, pH 8.0) is
20 required for the effective elution of the bound complex. The Zn-IMAC step separates the soluble OP-1 from the bulk of the contaminating serum proteins that elute in the flow through and 35 mM imidazole wash fractions. The Zn-IMAC purified soluble OP-1 is next
25 applied to an S-Sepharose cation-exchange column equilibrated in 20 mM NaPO_4 (pH 7.0) with 50 mM NaCl. This S-Sepharose step serves to further purify and concentrate the soluble OP-1 complex in preparation for the following gel filtration step. The protein was

- 35 -

applied to a Sephacryl S-200HR column equilibrated in TBS. Using substantially the same protocol, soluble morphogens also may be isolated from one or more body fluids, including serum, cerebro-spinal fluid or
5 peritoneal fluid.

IMAC was performed using Chelating-Sepharose (Pharmacia) that had been charged with three column volumes of 0.2 M ZnSO_4 . The conditioned media was
10 titrated to pH 7.0 and applied directly to the ZN-IMAC resin equilibrated in 20 mM HEPES (pH 7.0) with 500 mM NaCl. The Zn-IMAC resin was loaded with 80 mL of starting conditioned media per mL of resin. After loading the column was washed with equilibration buffer
15 and most of the contaminating proteins were eluted with 35 mM imidazole (pH 7.0) in equilibration buffer. The soluble OP-1 complex is then eluted with 50 mM imidazole (pH 8.0) in 20 mM HEPES and 500 mM NaCl.

20 The 50 mM imidazole eluate containing the soluble OP-1 complex was diluted with nine volumes of 20 mM NaPO_4 (pH 7.0) and applied to an S-Sepharose (Pharmacia) column equilibrated in 20 mM NaPO_4 (pH 7.0) with 50 mM NaCl. The S-Sepharose resin was loaded with
25 an equivalent of 800 mL of starting conditioned media per mL of resin. After loading the S-Sepharose column was washed with equilibration buffer and eluted with 100 mM NaCl followed by 300 mM and 500 mM NaCl in 20 mM NaPO_4 (pH 7.0). The 300 mM NaCl pool was further
30 purified using gel filtration chromatography. Fifty mls of the 300 mM NaCl eluate was applied to a 5.0 X 90 cm Sephacryl S-200HR (Pharmacia) equilibrated in Tris buffer d saline (TBS), 50 mM Tris, 150 mM NaCl (pH 7.4). The column was eluted at a flow rate of 5

- 36 -

mL/minute collecting 10 mL fractions. The apparent molecular of the soluble OP-1 was determined by comparison to protein molecular weight standards (alcohol dehydrogenase (ADH, 150 kDa), bovine serum albumin (BSA, 68 kDa), carbonic anhydrase (CA, 30 kDa) and cytochrome C (cyt C, 12.5 kDa). (see Fig. 3) The purity of the S-200 column fractions was determined by separation on standard 15% polyacrylamide SDS gels stained with coomassie blue. The identity of the mature OP-1 and the pro-domain was determined by N-terminal sequence analysis after separation of the mature OP-1 from the pro-domain using standard reverse phase C18 HPLC.

Figure 3 shows the absorbance profile at 280 nm. The soluble OP-1 complex elutes with an apparent molecular weight of 110 kDa. This agrees well with the predicted composition of the soluble OP-1 complex with one mature OP-1 dimer (35-36 kDa) associated with two pro-domains (39 kDa each). Purity of the final complex can be verified by running the appropriate fraction in a reduced 15% polyacrylamide gel.

The complex components can be verified by running the complex-containing fraction from the S-200 or S-200HR columns over a reverse phase C18 HPLC column and eluting in an acetonitrile gradient (in 0.1% TFA), using standard procedures. The complex is dissociated by this step, and the pro domain and mature species elute as separate species. These separate species then can be subjected to N-terminal sequencing using standard procedures (see, for example, Guide to Protein Purification, M. Deutscher, ed., Academic Press, San Diego, 1990, particularly pp. 602-613), and

- 37 -

the identity of the isolated 36kD, 39kDa proteins confirmed as mature morphogen and isolated, cleaved pro domain, respectively. N-terminal sequencing of the isolated pro domain from mammalian cell produced OP-1 revealed 2 forms of the pro region, the intact form (beginning at residue 30 of Seq. ID No. 1) and a truncated form, (beginning at residue 48 of Seq. ID No. 1.) N-terminal sequencing of the polypeptide subunit of the isolated mature species reveals a range of N-termini for the mature sequence, beginning at residues 293, 300, 313, 315, 316, and 318, of Seq. ID No. 1, all of which are active as demonstrated by the standard bone induction assay.

15 V. In Vitro Soluble Morphogen Complex Formation

As an alternative to purifying soluble complexes from culture media or a body fluid, soluble complexes may be formulated from purified pro domains and mature dimeric species. Successful complex formation apparently requires association of the components under denaturing conditions sufficient to relax the folded structure of these molecules, without affecting disulfide bonds. Preferably, the denaturing conditions mimic the environment of an intracellular vesicle sufficiently such that the cleaved pro domain has an opportunity to associate with the mature dimeric species under relaxed folding conditions. The concentration of denaturant in the solution then is decreased in a controlled, preferably step-wise manner, so as to allow proper refolding of the dimer and pro regions while maintaining the association of the pro

- 38 -

domain with the dimer. Useful denaturants include 4-6M urea or guanidine hydrochloride (GuHCl), in buffered solutions of pH 4-10, preferably pH 6-8. The soluble complex then is formed by controlled dialysis or
5 dilution into a solution having a final denaturant concentration of less than 0.1-2M urea or GuHCl, preferably 1-2 M urea or GuHCl, which then preferably can be diluted into a physiological buffer. Protein purification/renaturing procedures and considerations
10 are well described in the art, and details for developing a suitable renaturing protocol readily can be determined by one having ordinary skill in the art. One useful text on the subject is Guide to Protein Purification, M. Deutscher, ed., Academic Press, San
15 Diego, 1990, particularly section V. Complex formation also may be aided by addition of one or more chaperone proteins.

VI. Stability of Soluble Morphogen Complexes

20 The stability of the highly purified soluble morphogen complex in a physiological buffer, e.g., tris-buffered saline (TBS) and phosphate-buffered saline (PBS), can be enhanced by any of a number of
25 means. Currently preferred is by means of a pro region that comprises at least the first 18 amino acids of the pro sequence (e.g., residues 30-47 of Seq. ID NO. 1 for OP-1), and preferably is the full length pro region. Residues 30-47 show sequence homology to the N-terminal
30 portion of other morphogens and are believed to have particular utility in enhancing complex stability for

- 39 -

all morphogens. Other useful means for enhancing the stability of soluble morphogen complexes include three classes of additives. These additives include basic amino acids (e.g., L-arginine, lysine and betaine);
5 nonionic detergents (e.g., Tween 80 or Nonidet P-120); and carrier proteins (e.g., serum albumin and casein). Useful concentrations of these additives include 1-100 mM, preferably 10-70 mM, including 50 mM, basic amino acid;; 0.01-1.0%, preferably 0.05-0.2%, including 0.1%
10 (v/v) nonionic detergent;; and 0.01-1.0%, preferably 0.05-0.2%, including 0.1% (w/v) carrier protein.

VII. Activity of Soluble Morphogen Complex

15 Association of the pro domain with the mature dimeric species does not interfere with the morphogenic activity of the protein in vivo as demonstrated by different activity assays. Specifically, soluble OP-1 complex provided in a standard rat osteopenia model
20 induces significant increase in bone growth and osteocalcin production (see Table II, below), in a manner analogous to the results obtained using mature morphogen.

25 The assay is analogous to the osteoporosis model described in international application US92/07432 (WO93/05751), but uses aged female rats rather than ovariectomized animals. Briefly, young or aged female rats (Charles River Labs, 115-145, and 335-460g body
30 weight, respectively) were dosed daily for 7 days by intravenous tail injection, with either 20 µg/Kg body weight soluble OP-1, or 100 µg/Kg body weight soluble OP-1. Control groups of young and aged female rats were dosed only with tris-buffered saline (TBS). Water

- 40 -

and food were provided to all animals ad libitum. After 14 days, animals were sacrificed, and new bone growth measured by standard histometric procedures. Osteocalcin concentrations in serum also were measured.

5 No detrimental effects of morphogen administration were detected as determined by changes in animal body or organ weight or by hematology profiles.

TABLE II

| 10 | No. Animals | Animal Group | Bone Area (B.Ar/T.Ar) | Osteocalcin (ng/ml) |
|----|----------------|---|--------------------------|------------------------|
| 15 | 4 | Control | 5.50 \pm 0.64 | 11.89 \pm 4.20 |
| 20 | 5 | Aged female, 20 μ g/Kg sol. OP-1 | 7.68 \pm 0.63** | 22.24 \pm 2.28** |
| 25 | 5 | Aged female, 100 μ g/Kg sol. OP-1 | 9.82 \pm 3.31* | 20.87 \pm 6.14* |

*P < 0.05

**P < 0.01

30 Similar experiments performed using soluble OP-1 complex in the osteoporosis model described in WO93/05751 using ovariectomized rats also show no detrimental effect using the complex form.

35 Both mature and soluble morphogen also can induce CAM (cell adhesion molecule) expression, as demonstrated below. Briefly, induction of N-CAM isoforms (N-CAM-180, N-CAM-140 and N-CAM-120) can be monitored by reaction with the commercially available

40 antibody mAb H28.123 (Sigma Co., St. Louis) and

- 41 -

available antibody mAb H28.123 (Sigma Co., St. Louis) and standard Western blot analysis (see, for example, Molecular Cloning, A Laboratory Manual, Sambrook et al. eds. Cold Spring Harbor Press, New York, 1989, particularly Section 18). Incubation of a growing culture of transformed cells of neuronal origin, NG108-15 cells (ATCC, Rockville, MD) with either mature morphogen dimers or soluble morphogen complexes (10-100 ng/ml, preferably at least 40 ng/ml) induces a redifferentiation of these cells back to a morphology characteristic of untransformed neurons, including specific induction and/or enhanced expression of all 3 N-CAM isoforms. In the experiment, cells were subcultured on poly-L-lysine coated 6-well plates and grown in chemically defined medium for 2 days before the experiment. Fresh aliquots of morphogen were added (2.5 μ l) daily.

VIII. Antibody Production

20

Provided below are standard protocols for polyclonal and monoclonal antibody production. For antibodies which recognize the soluble complex only, preferably the isolated pro region is used as the antigen; where antibodies specific to the mature protein are desired, the antigen preferably comprises at least the C-terminal domain or the intact mature sequence.

30

Polyclonal antibody may be prepared as follows. Each rabbit is given a primary immunization of 100 μ g/500 μ l of antigen, in 0.1% SDS mixed with 500 μ l Complete Freund's Adjuvant. The antigen is injected

- 42 -

subcutaneously at multiple sites on the back and flanks of the animal. The rabbit is boosted after a month in the same manner using incomplete Freund's Adjuvant. Test bleeds are taken from the ear vein seven days
5 later. Two additional boosts and test bleeds are performed at monthly intervals until antibody against the morphogen antigen is detected in the serum using an ELISA assay. Then, the rabbit is boosted monthly with 100 μ g of antigen and bled (15 ml per bleed) at days
10 seven and ten after boosting.

Monoclonal antibody specific for a given morphogen may be prepared as follows. A mouse is given two injections of the morphogen antigen. The protein or
15 protein fragment preferably is recombinantly produced. The first injection contains 100 μ g of antigen in complete Freund's adjuvant and is given subcutaneously. The second injection contains 50 μ g of antigen in incomplete adjuvant and is given intraperitoneally.
20 The mouse then receives a total of 230 μ g of OP-3 in four intraperitoneal injections at various times over an eight month period. One week prior to fusion, the mouse is boosted intraperitoneally with antigen (e.g., 100 μ g) and may be additionally boosted with a peptide
25 fragment conjugated to bovine serum albumin with a suitable crosslinking agent. This boost can be repeated five days (IP), four days (IP), three days (IP) and one day (IV) prior to fusion. The mouse spleen cells then are fused to commercially available
30 myeloma cells at a ratio of 1:1 using PEG 1500

- 43 -

(Boeringer Mannheim, Germany), and the fused cells plated and screened for mature or soluble morphogen-specific antibodies using the appropriate portion of the morphogen sequence as antigen. The cell fusion and
5 monoclonal screening steps readily are performed according to standard procedures well described in standard texts widely available in the art.

Using these standard procedures, anti-pro domain
10 antisera was prepared from rabbits using the isolated pro domain from OP-1 as the antigen, and monoclonal antibody ("mAb") to the mature domain was produced in mice, using an E. coli-produced truncated form of OP-1 as antigen.

15
Standard Western blot analysis performed under reducing conditions demonstrates that the anti-pro domain antisera ("anti-pro") is specific for the pro domain only, while the mAb to mature OP-1 ("anti-mature
20 OP-1") is specific for the dimer subunits, that the two antibodies do not cross-react, and that the antibodies and can be used to distinguish between soluble and mature protein forms in a sample, e.g., of conditioned media or serum. A tabular representation of the
25 Western blot results is in Table III below, where reactivity of mAb to mature OP-1 is indicated by "yy", and reactivity of the anti-pro antisera is indicated by "xx".

- 44 -

TABLE III

| | <u>Antibody</u> | <u>Purified Sol OP1</u> | <u>Conditioned CHO Cell Media</u> | <u>Isolated Pro Domain</u> | <u>Purified Dimer Subunits</u> |
|----|------------------------|-----------------------------|---------------------------------------|--------------------------------|--|
| 5 | "anti-pro" | xx | xx | xx | |
| 10 | "anti- mature OP-1" | yy | yy | | yy |

15 IX. Immunoassays

20 The ability to detect morphogens in solution and to distinguish between soluble and mature dimeric morphogen forms provides a valuable tool for diagnostic assays, allowing one to monitor the level and type of morphogen free in the body, e.g., in serum and other body fluids, as well as to develop diagnostic and other tissue evaluating kits.

25 For example, OP-1 is an intimate participant in normal bone growth and resorption. Thus, soluble OP-1 is expected to be detected at higher concentrations in individuals experiencing high bone turnover, such as children, and at substantially lower levels in
30 individuals with abnormally low rates of bone turnover, such as patients with osteoporosis, osteosarcoma, Paget's disease and the like. Monitoring the level of OP-1, or other bone targeted morphogens such as BMP2 and BMP4, in serum thus provides a means for evaluating
35 the status of bone tissue in an individual, as well as a means for monitoring the efficacy of a treatment to regenerate damaged or lost bone tissue. Similarly,

- 45 -

monitoring the level of endogenous GDF-1, can provide diagnostic information on the health of nerve tissue, particularly brain tissue. Moreover, following this disclosure one can distinguish between the level of
5 soluble and mature forms in solution.

A currently preferred detection means for evaluating the level of morphogen in a body fluid comprises an immunoassay utilizing an antibody or other
10 suitable binding protein capable of reacting specifically with a morphogen and being detected as part of a complex with the morphogen. Immunoassays may be performed using standard techniques known in the art and antibodies raised against a morphogen and specific
15 for that morphogen. Antibodies which recognize a morphogen protein form of interest may be generated as described herein and these antibodies then used to monitor endogenous levels of protein in a body fluid, such as serum, whole blood or peritoneal fluid. To
20 monitor endogenous concentrations of soluble morphogen, the antibody chosen preferably has binding specificity for the soluble form e.g., has specificity for the pro domain. Such antibodies may be generated by using the pro domain or a portion thereof as the antigen,
25 essentially as described herein. A suitable pro domain for use as an antigen may be obtained by isolating the soluble complex and then separating the noncovalently associated pro domain from the mature domain using standard procedures, e.g., by passing the complex over
30 an HPLC column, as described above or by separation by gel electrophoresis. Alternatively, the pro form of the protein in its monomeric form may be used as the

- 46 -

antigen and the candidate antibodies screened by Western blot or other standard immunoassay for those which recognize the pro domain of the soluble form of the protein of interest, but not the mature form, also
5 as described above.

Monomeric pro forms can be obtained from cell lysates of CHO produced cells, or from prokaryotic expression of a DNA encoding the pro form, in for
10 example, E.coli. The pro form, which has an apparent molecular weight of about 50 kDa in mammalian cells, can then be isolated by HPLC and/or by gel electrophoresis, as described above.

15 In order to detect and/or quantitate the amount of morphogenic protein present in a solution, an immunoassay may be performed to detect the morphogen using a polyclonal or monoclonal antibody specific for that protein. Here, soluble and mature forms of the
20 morphogen also may be distinguished by using antibodies that discriminate between the two forms of the proteins as described above. Currently preferred assays include ELISAS and radioimmunoassays, including standard competitor assays useful for quantitating the morphogen
25 in a sample, where an unknown amount of sample morphogen is allowed to react with anti-morphogen antibody and this interaction is competed with a known amount of labeled antigen. The level of bound or free labeled antigen at equilibrium then is measured to
30 quantitate the amount of unlabeled antigen in solution, the amount of sample antigen being proportional to the amount of free labeled antigen. Exemplary protocols for these assays are provided below. However, as will be appreciated by those skilled in the art, variations

- 47 -

of these protocols, as well as other immunoassays, are well known in the literature and within the skill of the art. For example, in the ELISA protocol provided below, soluble OP-1 is identified in a sample using

5 biotinylated anti-pro antiserum. Biotinylated antibodies can be visualized in a colormetric assay or in a chemiluminescent assay, as described below. Alternatively, the antibody can be radio-labeled with a suitable molecule, such as ^{125}I . Still another

10 protocol that may be used is a solid phase immunoassay, preferably using an affinity column with anti-morphogen antibody complexed to the matrix surface and over which a serum sample may be passed. A detailed description of useful immunoassays, including protocols and general

15 considerations is provided in, for example, Molecular Cloning: A Laboratory Manual, Sambrook et al., eds. Cold Spring Harbor Press, New York, 1989, particularly Section 18.

20 For serum assays, the serum preferably first is partially purified to remove some of the excess, contaminating serum proteins, such as serum albumin. Preferably the serum is extracted by precipitation in ammonium sulfate (e.g., 45%) such that the complex is

25 precipitated. Further purification can be achieved using purification strategies that take advantage of the differential solubility of soluble morphogen complex or mature morphogens relative to that of the other proteins present in serum. Further purification

30 also can be achieved by chromatographic techniques well known in the art.

- 48 -

Soluble OP-1 may be detected using a polyclonal antibody specific for the OP-1 pro domain in an ELISA, as follows. 1 μ g/100 μ l of affinity-purified polyclonal rabbit IgG specific for OP-1-pro is added to each well of a 96-well plate and incubated at 37°C for an hour. The wells are washed four times with 0.167M sodium borate buffer with 0.15 M NaCl (BSB), pH 8.2, containing 0.1% Tween 20. To minimize non-specific binding, the wells are blocked by filling completely with 1% bovine serum albumin (BSA) in BSB and incubating for 1 hour at 37°C. The wells are then washed four times with BSB containing 0.1% Tween 20. A 100 μ l aliquot of an appropriate dilution of each of the test samples of cell culture supernatant or serum sample is added to each well in triplicate and incubated at 37°C for 30 min. After incubation, 100 μ l biotinylated rabbit anti-pro serum (stock solution is about 1 mg/ml and diluted 1:400 in BSB containing 1% BSA before use) is added to each well and incubated at 37°C for 30 min. The wells are then washed four times with BSB containing 0.1% Tween 20. 100 μ l streptavidin-alkaline (Southern Biotechnology Associates, Inc. Birmingham, Alabama, diluted 1:2000 in BSB containing 0.1% Tween 20 before use) is added to each well and incubated at 37°C for 30 min. The plates are washed four times with 0.5M Tris buffered Saline (TBS), pH 7.2. 50 μ l substrate (ELISA Amplification System Kit, Life Technologies, Inc., Bethesda, MD) is added to each well incubated at room temperature for 15 min. Then, 50 μ l amplifier (from the same amplification system kit) is added and incubated for another 15 min at room temperature. The reaction is stopped by the addition of 50 μ l 0.3 M sulphuric acid.

- 49 -

The OD at 490 nm of the solution in each well is recorded. To quantitate the level of soluble OP-1 in the sample, a standard curve is performed in parallel with the test samples. In the standard curve, known
5 increasing amounts of purified OP-1-pro is added. Alternatively, using, for example, Lumi-phos 530 (Analytical Luminescence Laboratories) as the substrate and detection at 300-650 nm in a standard luminometer, complexes can be detected by chemiluminescence, which
10 typically provides a more sensitive assay than detection by means of a visible color change.

Morphogen (soluble or mature form) may be detected in a standard plated-based radioimmunoassay as follows.
15 Empirically determined limiting levels of anti-morphogen antibody (e.g., anti-OP-1, typically 50-80 ng/well) are bound to wells of a PVC plate e.g., in 50 μ l PBS phosphate buffered saline. After sufficient incubation to allow binding at room
20 temperature, typically one hour, the plate is washed in a PBS/Tween 20 solution, ("washing buffer"), and 200 μ l of block (3% BSA, 0.1 μ lysine in 1xBSB) is added to each well and allowed to incubate for 1 hour, after which the wells are washed again in washing buffer. 40
25 μ l of a sample composed of serially diluted plasma (preferably partially purified as described above) or morphogen standard (e.g., OP-1) is added to wells in triplicate. Samples preferably are diluted in PTTH (15 mM KH_2PO_4 , 8 mM Na_2PO_4 , 27 mM KCl, 137 mM NaCl,
30 0.05% Tween 20, 1 mg/ml HSA, 0.05% NaN_3 , pH 7.2). 10 μ l of labelled competitor antigen, preferably 100,000-500,000 cpm/sample is added (e.g., ^{125}I OP-1, radiolabelled using standard procedures), and plates are incubated overnight at 4°C. Plates then are washed

- 50 -

in washing buffer, and allowed to dry. Wells are cut apart and bound labelled OP-1 counted in a standard gamma counter. The quantities of bound labelled antigen (e.g., ^{125}I OP-1) measured in the presence and
5 absence of sample then are compared, the difference being proportional to the amount of sample antigen (morphogen) present in the sample fluid.

As a corollary assay method, immunoassays may be
10 developed to detect endogenous anti-morphogen antibodies, and to distinguish between such antibodies to soluble or mature forms. Endogenous anti-morphogen antibodies have been detected in serum, and their level is known to increase, for example, upon implanting of
15 an osteogenic device in a mammal. Without being limited to a particular theory, these antibodies may play a role in modulating morphogen activity by modulating the level of available protein in serum. Assays that monitor the level of endogenous antibodies
20 in blood or their body fluids thus can be used in diagnostic assays to evaluate the status of a tissue, as well as to provide a means for monitoring the efficacy of a therapy for tissue regeneration.

25 The currently preferred means for detecting endogenous anti-morphogen antibodies is by means of a standard Western blot. See, for example, Molecular Cloning: A Laboratory Manual Sambrook et al., eds., Cold Spring Harbor Press, New York, 1989, particularly
30 pages 18.60-18.75, incorporated herein by reference, for a detailed description of these assays. Purified mature or soluble morphogen is electrophoresed on an SDS polyacrylamide gel under oxidized or reduced conditions designed to separate the proteins in

- 51 -

solution, and the proteins then transferred to a polyvinylidene difluoride microporus membrane (0.45 μ m pore sizes) using standard buffers and procedures. The filter then is incubated with the
5 serum being tested (at various dilutions). Antibodies bound to either the pro domain or the mature morphogen domain are detected by means of an anti-human antibody protein, e.g., goat anti-human Ig. Titers of the antimorphogen antibodies can be determined by further
10 dilution of the serum until no signal is detected.

X. Formulations and Methods for Administering Soluble Morphogens as Therapeutic Agents

15 The soluble morphogens of this invention are particularly useful as therapeutic agents to regenerate diseased or damaged tissue in a mammal, particularly a human.

20 The soluble morphogen complexes may be used to particular advantage in regeneration of damaged or diseased lung, heart, liver, kidney, nerve or pancreas tissue, as well as in the transplantation and/or grafting of these tissues and bone marrow, skin,
25 gastrointestinal mucosa, and other living tissues.

The soluble morphogen complexes described herein may be provided to an individual by any suitable means, preferably directly or systemically, e.g., parenterally
30 or orally. Where the morphogen is to be provided directly (e.g., locally, as by injection, to a desired tissue site), or parenterally, such as by intravenous, subcutaneous, intramuscular, intraorbital, ophthalmic, intraventricular, intracranial, intracapsular,

- 52 -

intraspinal, intracisternal, intraperitoneal, buccal, rectal, vaginal, intranasal or by aerosol administration, the soluble morphogen complex preferably comprises part of an aqueous solution. The
5 solution is physiologically acceptable so that in addition to delivery of the desired morphogen to the patient, the solution does not otherwise adversely affect the patient's electrolyte and volume balance. The aqueous medium for the soluble morphogen thus may
10 comprise normal physiologic saline (0.9% NaCl, 0.15M), pH 7-7.4.

Soluble morphogens of this invention are readily purified from cultured cell media into a physiological
15 buffer, as described above. In addition, and as described above, if desired, the soluble complexes may be formulated with one or more additional additives, including basic amino acids (e.g., L-arginine, lysine, betaine); non-ionic detergents (e.g. Tween-80 or
20 Nonidet-120) and carrier proteins (e.g., serum albumin and casein).

Useful solutions for oral or parenteral administration may be prepared by any of the methods
25 well known in the pharmaceutical art, described, for example, in Remington's Pharmaceutical Sciences, (Gennaro, A., ed.), Mack Pub., 1990. Formulations may include, for example, polyalkylene glycols such as polyethylene glycol, oils of vegetable origin,
30 hydrogenated naphthalenes, and the like. Formulations for direct administration, in particular, may include glycerol and other compositions of high viscosity.

- 53 -

Biocompatible, preferably bioresorbable polymers, including, for example, hyaluronic acid, collagen, tricalcium phosphate, polybutyrate, polylactide, polyglycolide and lactide/glycolide copolymers, may be
5 useful excipients to control the release of the soluble morphogen in vivo.

Other potentially useful parenteral delivery systems for these morphogens include ethylene-vinyl
10 acetate copolymer particles, osmotic pumps, implantable infusion systems, and liposomes. Formulations for inhalation administration may contain as excipients, for example, lactose, or may be aqueous solutions containing, for example, polyoxyethylene-9-lauryl
15 ether, glycocholate and deoxycholate, or oily solutions for administration in the form of nasal drops, or as a gel to be applied intranasally.

The soluble morphogens described herein also may be
20 administered orally. Oral administration of proteins as therapeutics generally is not practiced as most proteins readily are degraded by digestive enzymes and acids in the mammalian digestive system before they can be absorbed into the bloodstream. However, the mature
25 domains of the morphogens described herein typically are acid-stable and protease-resistant (see, for example, U.S. Pat. No. 4,968,590.) In addition, at least one morphogen, OP-1, has been identified, in mammary gland extract, colostrum and milk, as well as
30 saliva. Moreover, the OP-1 purified from mammary gland extract is morphogenically active. For example, this protein induces endochondral bone formation in mammals when implanted subcutaneously in association with a suitable matrix material, using a standard in vivo bone

- 54 -

assay, such as is disclosed in U.S. Pat. No. 4,968,590. In addition, endogenous morphogen also is detected in human serum (see above). Finally, comparative experiments with soluble and mature morphogens in a
5 number of experiments defining morphogenic activity indicate that the non-covalent association of the pro domain with the dimeric species does not interfere with morphogenic activity. These findings indicate that oral and parenteral administration are viable means for
10 administering morphogens to an individual, and that soluble morphogens have utility in systemic administration protocols.

The soluble complexes provided herein also may be
15 associated with molecules capable of targeting the morphogen to a desired tissue. For example, tetracycline and diphosphonates (bisphosphonates) are known to bind to bone mineral, particularly at zones of bone remodeling, when they are provided systemically in
20 a mammal. Accordingly, these molecules may be included as useful agents for targeting soluble morphogens to bone tissue. Alternatively, an antibody or other binding protein that interacts specifically with a surface molecule on the desired target tissue cells
25 also may be used. Such targeting molecules further may be covalently associated to the morphogen complex, e.g., by chemical crosslinking, or by using standard genetic engineering means to create, for example, an acid labile bond such as an Asp-Pro linkage. Useful
30 targeting molecules may be designed, for example, using the single chain binding site technology disclosed, for example, in U.S. Pat. No. 5,091,513.

- 55 -

Finally, the soluble morphogen complexes provided herein may be administered alone or in combination with other molecules known to have a beneficial effect on tissue morphogenesis, including molecules capable of
5 tissue repair and regeneration and/or inhibiting inflammation. Examples of useful cofactors for stimulating bone tissue growth in osteoporotic individuals, for example, include but are not limited to, vitamin D₃, calcitonin, prostaglandins, parathyroid
10 hormone, dexamethasone, estrogen and IGF-I or IGF-II. Useful cofactors for nerve tissue repair and regeneration may include nerve growth factors. Other useful cofactors include symptom-alleviating cofactors, including antiseptics, antibiotics, antiviral and
15 antifungal agents and analgesics and anesthetics.

The compounds provided herein can be formulated into pharmaceutical compositions by admixture with pharmaceutically acceptable nontoxic excipients and
20 carriers. As noted above, such compositions may be prepared for parenteral administration, particularly in the form of liquid solutions or suspensions; for oral administration, particularly in the form of tablets or capsules; or intranasally, particularly in the form of
25 powders, nasal drops or aerosols. Where adhesion to a tissue surface is desired the composition may include the morphogen dispersed in a fibrinogen-thrombin composition or other bioadhesive such as is disclosed, for example in PCT US91/09275, the disclosure of which
30 is incorporated herein by reference. The composition then may be painted, sprayed or otherwise applied to the desired tissue surface.

- 56 -

The compositions can be formulated for parenteral or oral administration to humans or other mammals in therapeutically effective amounts, e.g., amounts which provide appropriate concentrations of the morphogen to target tissue for a time sufficient to induce morphogenesis, including particular steps thereof, as described above.

Where the soluble morphogen complex is to be used as part of a transplant procedure, the morphogen may be provided to the living tissue or organ to be transplanted prior to removal of the tissue or organ from the donor. The morphogen may be provided to the donor host directly, as by injection of a formulation comprising the soluble complex into the tissue, or indirectly, e.g., by oral or parenteral administration, using any of the means described above.

Alternatively or, in addition, once removed from the donor, the organ or living tissue may be placed in a preservation solution containing the morphogen. In addition, the recipient also preferably is provided with the morphogen just prior to, or concomitant with, transplantation. In all cases, the soluble complex may be administered directly to the tissue at risk, as by injection to the tissue, or it may be provided systemically, either by oral or parenteral administration, using any of the methods and formulations described herein and/or known in the art.

Where the morphogen comprises part of a tissue or organ preservation solution, any commercially available preservation solution may be used to advantage. A

- 57 -

useful preservation solution is described in in PCT/US92/07358 (WO93/04692), incorporated herein by reference.

5 As will be appreciated by those skilled in the art, the concentration of the compounds described in a therapeutic composition will vary depending upon a number of factors, including the dosage of the drug to be administered, the chemical characteristics (e.g.,
10 hydrophobicity) of the compounds employed, and the route of administration. The preferred dosage of drug to be administered also is likely to depend on such variables as the type and extent of tissue loss or defect, the overall health status of the particular
15 patient, the relative biological efficacy of the compound selected, the formulation of the compound, the presence and types of excipients in the formulation, and the route of administration. In general terms, the compounds of this invention may be provided in an
20 aqueous physiological buffer solution containing about 0.001 to 10% w/v compound for parenteral administration. Typical dose ranges are from about 10 ng/kg to about 1 g/kg of body weight per day; a preferred dose range is from about 0.1 μ g/kg to
25 100 mg/kg of body weight. No obvious morphogen-induced pathological lesions are induced when mature morphogen (e.g., OP-1, 20 μ g) is administered daily to normal growing rats for 21 consecutive days. Moreover, 10 μ g systemic injections of morphogen (e.g., OP-1) injected
30 daily for 10 days into normal newborn mice does not produce any gross abnormalities.

- 58 -

Where morphogens are administered systemically, in the methods of the present invention, preferably a large volume loading dose is used at the start of the treatment. The treatment then is continued with a
5 maintenance dose. Further administration then can be determined by monitoring at intervals the levels of the morphogen in the blood.

Other Embodiments

10

The invention may be embodied in other specific forms without departing from the spirit or essential characteristics thereof. The present embodiments are therefore to be considered in all respects as
15 illustrative and not restrictive, the scope of the invention being indicated by the appended claims rather than by the foregoing description, and all changes which come within the meaning and range of equivalency of the claims are therefore intended to be embraced
20 therein.

- 107 -

What is claimed is:

1. Dimeric protein comprising a pair of protein subunits associated to defined a dimeric structure having morphogenic activity,
each of said subunits comprising at least a 100 amino acid sequence having a pattern of cysteine residues characteristic of the morphogen family,
at least one of said subunits comprising a mature form of a subunit of a member of the morphogen family, or an allelic, species, or sequence variant thereof, noncovalently complexed with
a peptide comprising a pro region of a member of the morphogen family, or an allelic, species, or sequence variant thereof to form a complex which is more soluble in aqueous solvents than the uncomplexed pair of subunits.
2. The protein of claim 1 wherein both said subunits comprise a mature form of a subunit of a member of the morphogen family or an allelic, species, or sequence variant thereof, each said subunit being noncovalently complexed with a said peptide.
3. The protein of claim 1 wherein each said subunit is the mature form of human OP-1, or a species or allelic variant thereof.
4. The protein of claim 1, 2, or 3 wherein the peptide comprises the pro region of human OP-1, or a species, allelic or sequence variant thereof.

- 108 -

5. The protein of claim 1 wherein said peptide comprises at least the first 18 amino acids of an amino acid sequence defining said pro region.

6. The protein of claim 1 wherein said peptide comprises at least the first 18 amino acids of an amino acid sequence defining said pro region in Seq. ID Nos. 1-16 or a sequence variant thereof.

7. The protein of claim 1 or 6 wherein said peptide comprises the full length form of said pro region.

8. The protein of claim 1 wherein said pro region peptide comprises an amino acid sequence selected from sequences defined by residues 30-48, 30-292 and 48-292 of Seq. ID No. 1.

9. The protein of claim 1 wherein said pro region peptide comprises an amino acid sequence encoded by a nucleic acid that hybridizes under stringent conditions with a DNA encoding the N-terminal 18 amino acids of the pro region sequences for Seq. ID Nos. 1-19.

10. The protein of claims 1 or 9 wherein said pro region peptide comprises a nucleic acid that hybridizes under stringent conditions with a DNA defined by nucleotides of 136-192 of Seq. ID No. 1 or nucleotides 157-211 of Seq. ID No. 5.

11. The protein of claim 1 wherein said subunit sequence variant comprises a chimeric morphogen amino acid sequence.

- 109 -

12. The protein of claim 1 wherein said peptide comprises a chimeric pro region amino acid sequence.

13. The protein of claim 1 wherein said subunit comprises a sequence defined by Generic Sequence 7 or Generic Sequence 8.

14. The protein of claim 1 wherein said subunit comprises a sequence having 60% amino acid identity with the sequence defined by residues 335-431 of Seq. ID No.1.

15. The protein of claim 1 wherein said subunit comprises the mature form of a subunit defined by any of the sequences of Seq. ID No. 5-19.

16. The protein of claim 1 wherein said subunit comprises an amino acid sequence encoded by a nucleic acid that hybridizes with a DNA defined by nucleotides 1036-1341 of Seq. ID No. 1 or nucleotides 1390-1695 of Seq. ID No. 5.

17. The protein of claim 1 further comprising an molecule capable of enhancing the stability of said complex.

18. A therapeutic composition comprising the protein of any of claims 1, 2, 5-9 or 11-17.

19. A therapeutic composition comprising the protein of claim 1 wherein each said subunit is the mature form of human OP-1, or a species or allelic variant thereof.

- 110 -

20. A therapeutic composition comprising the protein of claim 1, wherein said peptide comprises part or all of the pro region of human OP-1, or a species or allelic variant thereof.
21. The therapeutic composition of claim 18 comprising the protein of claim 1 wherein said subunit comprises the mature form of a subunit defined by any of the sequences of Seq. ID Nos. 5-19.
22. A therapeutic composition comprising the protein of claims 3, 4 or 10.
23. The therapeutic composition of claims 18 or 22 further comprising a cofactor.
24. The therapeutic composition of claim 23 wherein said cofactor is a symptom-alleviating cofactor.
25. A kit for diagnosing a tissue disorder or evaluating the efficacy of a therapy to regenerate lost or damaged tissue in a mammal, the kit comprising:
- a) means for capturing a cell or fluid sample from said mammal,
 - b) a binding protein capable of interacting specifically with a soluble morphogen complex in said sample, and
 - c) means for detecting the binding protein bound to said soluble morphogen complex.
26. The kit of claim 25 wherein said binding protein is an antibody.

- 111 -

27. A method for evaluating the status of a tissue, the method comprising the step of comparing the quantity of morphogen in a body fluid sample with the quantity of morphogen in a control sample.

28. A method for evaluating the efficacy of a therapy to regenerate lost or damaged tissue in a mammal, the method comprising the step of comparing the quantity of morphogen in a body fluid sample with the quantity of morphogen in a control sample.

29. A method for diagnosing a tissue disorder in a mammal, the method comprising the step of comparing the quantity of morphogen in a body fluid sample with the quantity of morphogen in a control sample.

30. The invention of claim 25, 26, 27 or 28 wherein said morphogen is a dimeric protein comprising a pair of protein subunits associated to defined a dimeric structure having morphogenic activity,

each of said subunits comprising at least a 100 amino acid sequence having a pattern of cysteine residues characteristic of the morphogen family,

at least one of said subunits comprising a mature form of a subunit of a member of the morphogen family, or an allelic, species, or sequence variant thereof, noncovalently complexed with

a peptide comprising a pro region of a member of the morphogen family, or an allelic, species, or sequence variant thereof to form a complex which is more soluble in aqueous solvents than the uncomplexed pair of subunits.

- 112 -

31. The invention of claims 25, 26, 27 or 28 wherein said quantity of morphogen is detected by an immunoassay.

32. The invention of claims 25, 26, 27 or 28 wherein said quantity of morphogen is detected by an antibody capable of distinguishing soluble morphogen in a sample fluid.

33. The invention of claims 25, 26, 27 or 28 wherein said body fluid sample comprises serum.

34. The invention of claims 25 or 28 wherein said tissue disorder is a bone tissue disorder.

35. The invention of claim 34 wherein said bone tissue disorder is selected from the group consisting of osteosarcoma, osteoporosis, and Paget's disease.

36. A method of evaluating the status of a tissue, the method comprising the step of detecting the presence of anti-morphogen antibody in a tissue or body fluid sample.

37. A method for evaluating the efficacy of a therapy to regenerate lost or damaged tissue, the method comprising the step of detecting the presence of anti-morphogen antibody in a tissue or body fluid sample.

38. A method for diagnosing a tissue disorder, the method comprising the step of detecting the presence of anti-morphogen antibody in a tissue or body fluid sample.

- 113 -

39. A kit for diagnosing a tissue disorder or evaluating the efficacy of a therapy to regenerate lost or damaged tissue in a mammal, the kit comprising:

- a) means for capturing a cell or fluid sample from said mammal;
- b) a binding protein capable of interacting specifically with an endogenous anti-morphogen antibody in said sample; and
- c) means for detecting said binding protein-bound to said endogenous anti-morphogen antibody.

1/2

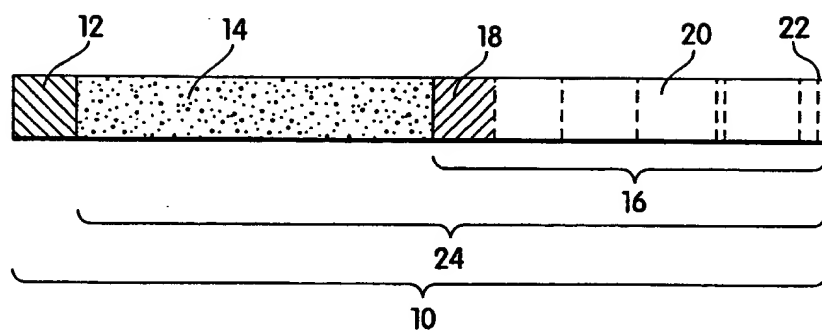


Fig. 1

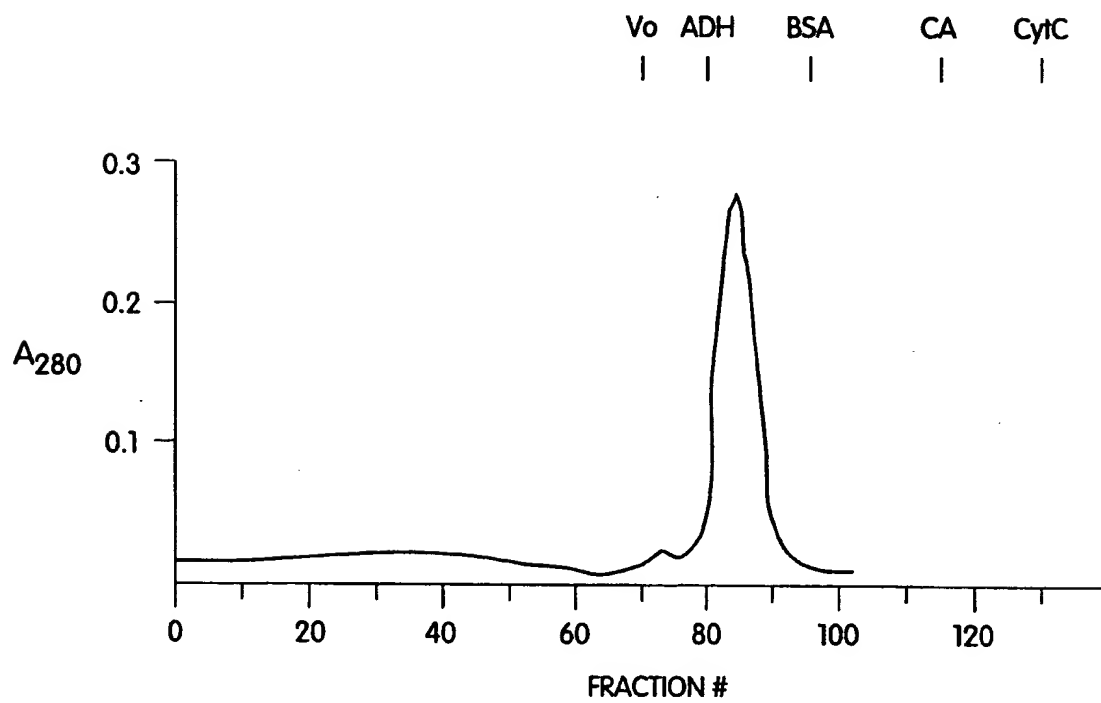


Fig. 3

SUBSTITUTE SHEET

2/2

OP-2: RAPRSQQPFVVTFFRASPSPIRTPRAVRPLRRRQPKKSNELPQANRLPGIFDDVHGSHGRQVC
 OP-1: RSIRSTGSKORSQNRSKTPKNQEALRMANVAENSSSDQRQAC
 Vgr-1: RTTRSASSRRRQQSRNRSTQSQDVSRGSSDYNGSELKTAC
 BMP-5: RSVRAANKRKNQNRNKSSSHQDSSRMSSVGDYNTSEQKQAC
 60A: RSKRSASHPRKRKKSVSPNNVPLLEPMESTRSC
 DPP: RSIRDVSGGEGGKGGRNKRHARRPTRRKNHDDTC
 BMP-2: RHVRISRSLHQDEHSWSQIRPLLVTFGHDGKGHPLHK--REKRQAKH--KQKRRLKSSC
 BMP-4: RISRSLPQGSNGNWAQLRPLLVTFGHDGRGHALTRRRRAKRSPKHHSORARKKNKNC
 Vg-1: RCKRPRRKRSYSKLPFTASNIC
 BMP-3: RKKRSTGVLLPLQ.....KSKNKKKQKGPHRKSTLQFDEQTLKKARRKQWIEPRNC

Fig. 2

INTERNATIONAL SEARCH REPORT

Int. Application No

PCT/US 93/07189

A. CLASSIFICATION OF SUBJECT MATTER

IPC 5 C12N15/12 A61K37/02 G01N33/50 G01N33/53

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 5 C07K C12N A61K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category * | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|------------|---|-----------------------|
| X | WO,A,91 18047 (GENENTECH, INC.) 28 November 1991 see page 2, line 24 - page 3, line 4 see page 4, line 4 - line 8 see page 5, line 16 - page 6, line 5 --- | 1,5,7,12 |
| X | MOLECULAR ENDOCRINOLOGY vol. 5, no. 1, January 1991 pages 149 - 155 R. GLENN HAMMONDS, JR. ET AL. 'Bone-inducing activity of mature BMP-2b produced from a hybrid BMP-2a/2b precursor' see abstract see page 149, right column, paragraph 3 - page 150, left column, paragraph 3 see page 152, left column, paragraph 2 - right column, paragraph 3 --- | 1,5,7,12 |
| -/-- | | |

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

* Special categories of cited documents:

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- "&" document member of the same patent family

Date of the actual completion of the international search

2 November 1993

Date of mailing of the international search report

14. 12. 93

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+ 31-70) 340-3016

Authorized officer

MONTERO LOPEZ, B

INTERNATIONAL SEARCH REPORT

Inter. Application No

PCT/US 93/07189

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

| Category * | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|------------|---|-----------------------|
| X | US,A,4 857 456 (MARSHALL R. URIST) 15 August 1989 cited in the application see column 1, line 12 - line 20 see column 2, line 37 - line 40; examples I-III ---- | 27-29, 36-39 |
| A | WO,A,92 07073 (CREATIVE BIOMOLECULES, INC.) 30 April 1992 see page 6, line 2 - line 8 see page 7, line 4 - page 9, line 9 ---- | 1-16 |
| P,X | WO,A,93 05751 (CREATIVE BIOMOLECULES, INC.) 1 April 1993 see page 7, line 16 - line 33 see page 10, line 5 - line 28 see page 11, line 6 - page 32, line 3 see page 37, line 17 - line 35 ----- | 1-24 |

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 93/07189

| Patent document cited in search report | Publication date | Patent family member(s) | Publication date |
|---|---------------------|---|--|
| WO-A-9118047 | 28-11-91 | US-A- 5168050 EP-A- 0531448 | 01-12-92 17-03-93 |
| US-A-4857456 | 15-08-89 | NONE | |
| WO-A-9207073 | 30-04-92 | AU-A- 8900091 CA-A- 2094027 | 20-05-92 19-04-92 |
| WO-A-9305751 | 01-04-93 | AU-A- 2564592 AU-A- 3176293 WO-A- 9304692 AU-A- 2862492 WO-A- 9305172 | 05-04-93 27-04-93 18-03-93 05-04-93 18-03-93 |